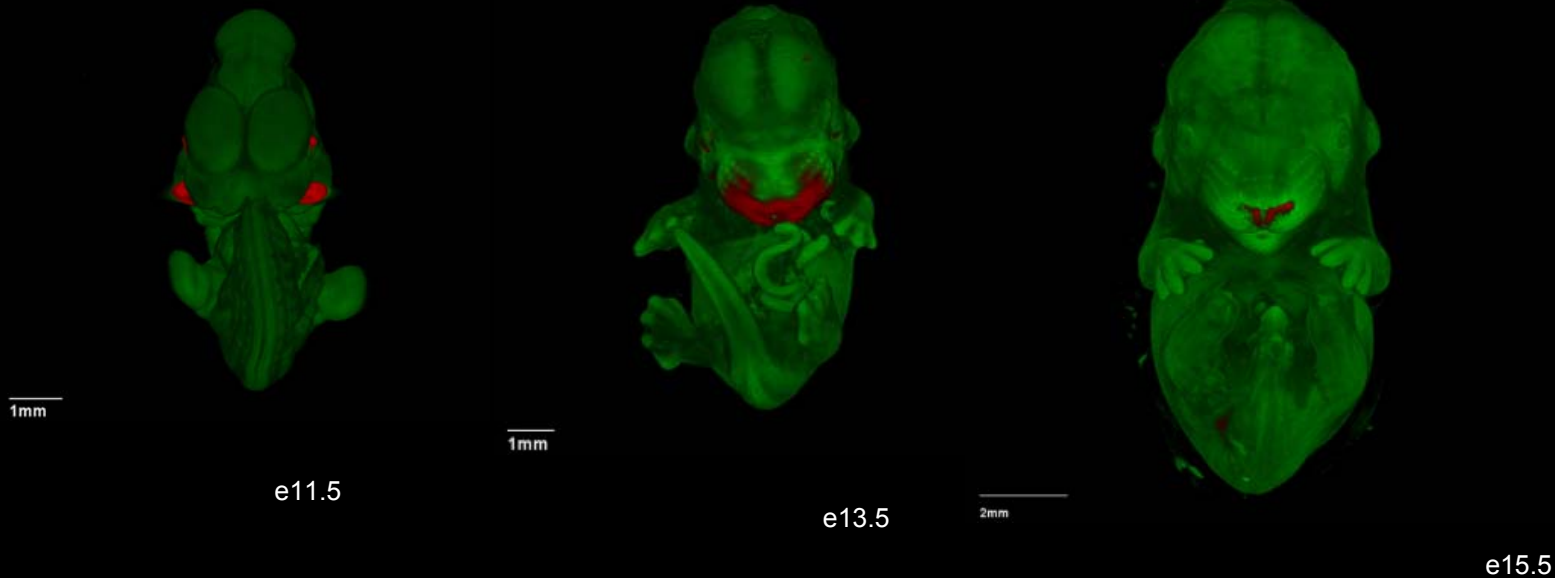


# Genomic and Transgenic Resources for Craniofacial Enhancer Studies

May 2018 Update



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Poster

sc RNA-Seq



**Sudha  
Rajderkar**

ChIP-/RNA-seq



**Cailyn  
Spurrell**

Informatics



**Matthew  
Blow**



**Valentina  
Snetkova**

# Outline

## Data Generation Progress

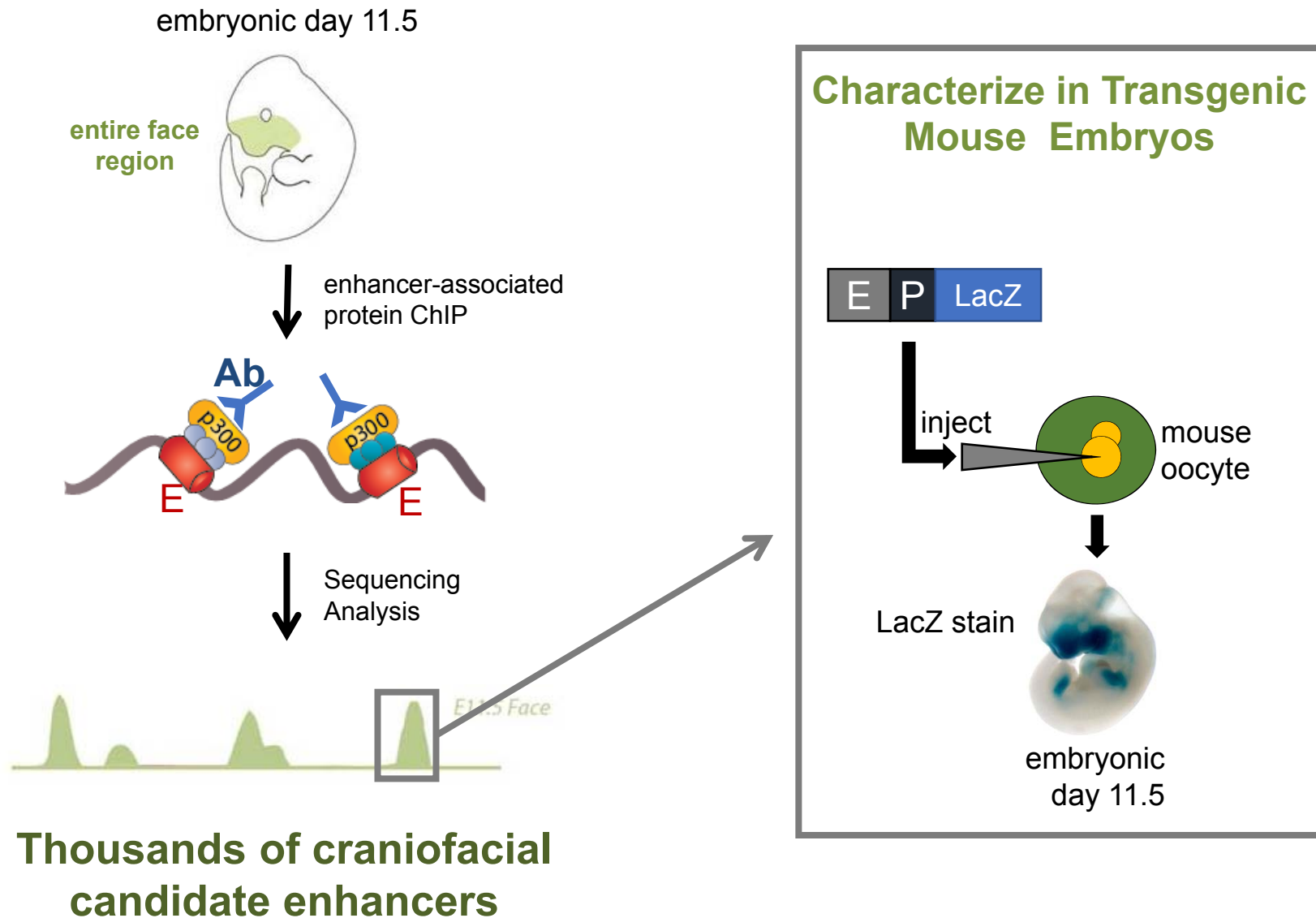
Chromatin and RNA-seq data from **human** and **mouse** craniofacial tissue, transgenic validation and OPT

## Analysis, Interactions and Collaborations

## Mapping Enhancers at Single Cell Resolution

## Centralized FaceBase Bioinformatics Pipeline

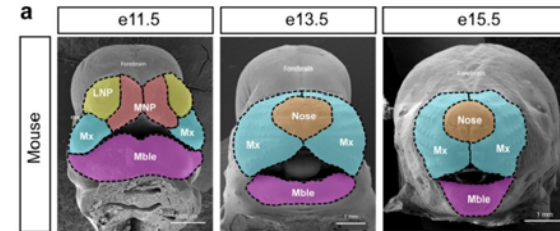
# Enhancer Mapping by Tissue-ChIP-seq



# Data Generation Progress

# FACEBASE 2: Specific Aims

## Aim 1a: Progress Mouse Tissues



### Mouse Tissues

Stage	Tissue	RNA-seq	H3K4me1	H3K27ac	H3K27me3	ATAC-seq
E11.5	Mandibular process	✓	✓	✓	✓	✓
	Maxillary process	✓	✓	✓	✓	✓
	Lateral nasal prominence	✓	✓	✓	✓	✓
	Medial nasal prominence	✓	✓	✓	✓	✓

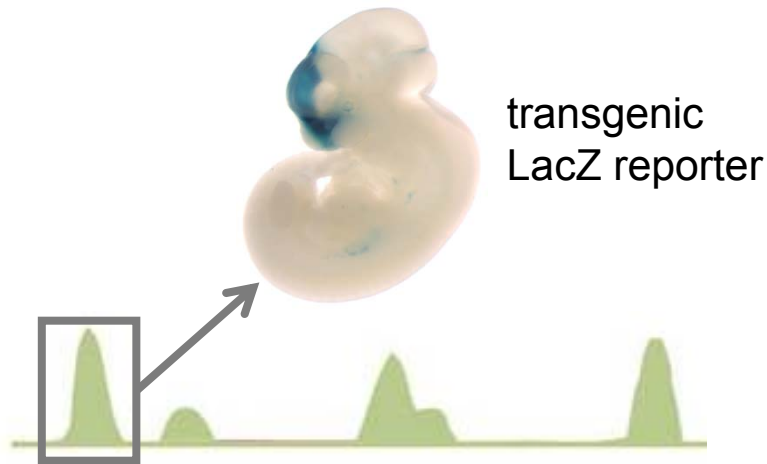
Stage	Tissue	RNA-seq	H3K4me1	H3K27ac	H3K27me3
E13.5	Mandibular process	✓	✓	✓	✓
	Maxillary process	✓	✓	✓	✓
	Nose	✓	✓	✓	✓

Stage	Tissue	RNA-seq	H3K4me1	H3K27ac	H3K27me3
E15.5	Mandibular process	✓	✓	✓	✓
	Maxillary process	✓	✓	✓	✓
	Nose	✓	✓	✓	✓

- major milestones complete
- in progress: replicates for E15.5

# FACEBASE 2: Specific Aims

## Aim 2: Transgenic Assays of Candidate Enhancer Sequences



- Candidate sequences:
  - Candidate regions from developmental mouse studies
  - Risk variants/alleles from human studies of CF birth defects (GWAS and Mendelian)
  - Variants linked to normal variation in GWAS
- 20 transgenic experiments in 2017
- 45% positive for craniofacial structures

**We make this capability available to other FaceBase investigators and are looking forward to collaborating!**

# FACEBASE 2: Specific Aims

**Aim 2: Optical projection tomography data (OPT)**  
**enhancer: background**

43 available in FaceBase Hub  
74 more generated to date

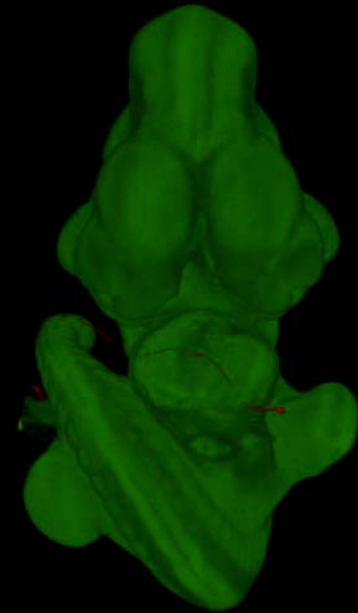


800 μm

hs59

Nasal process  
Branchial arch

Intragenic to *RPGRIP1L*  
Near *IRX3/4*



400 μm

hs921

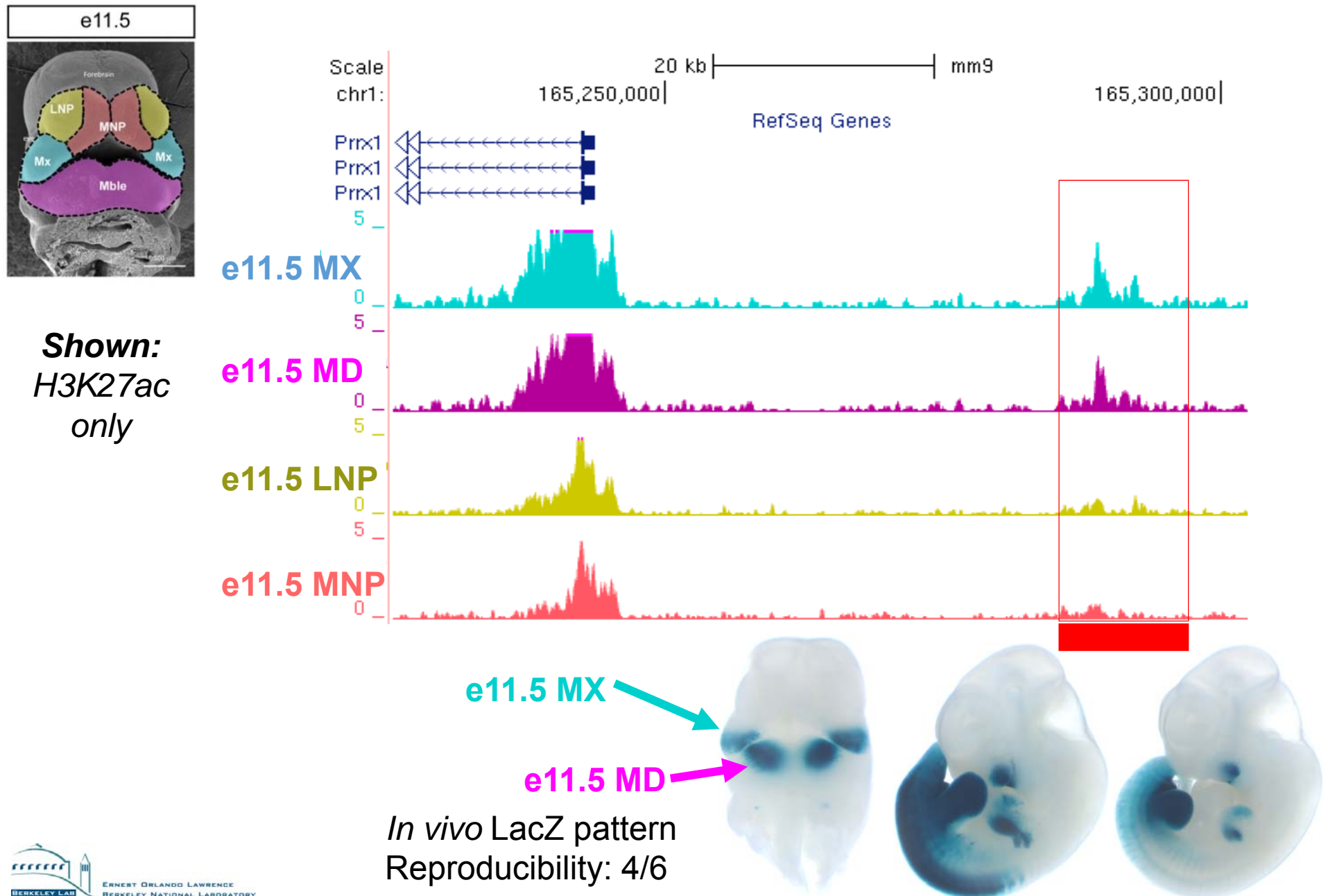
Mandibular process

Intragenic to *AGAP1*  
Near *GBX2*

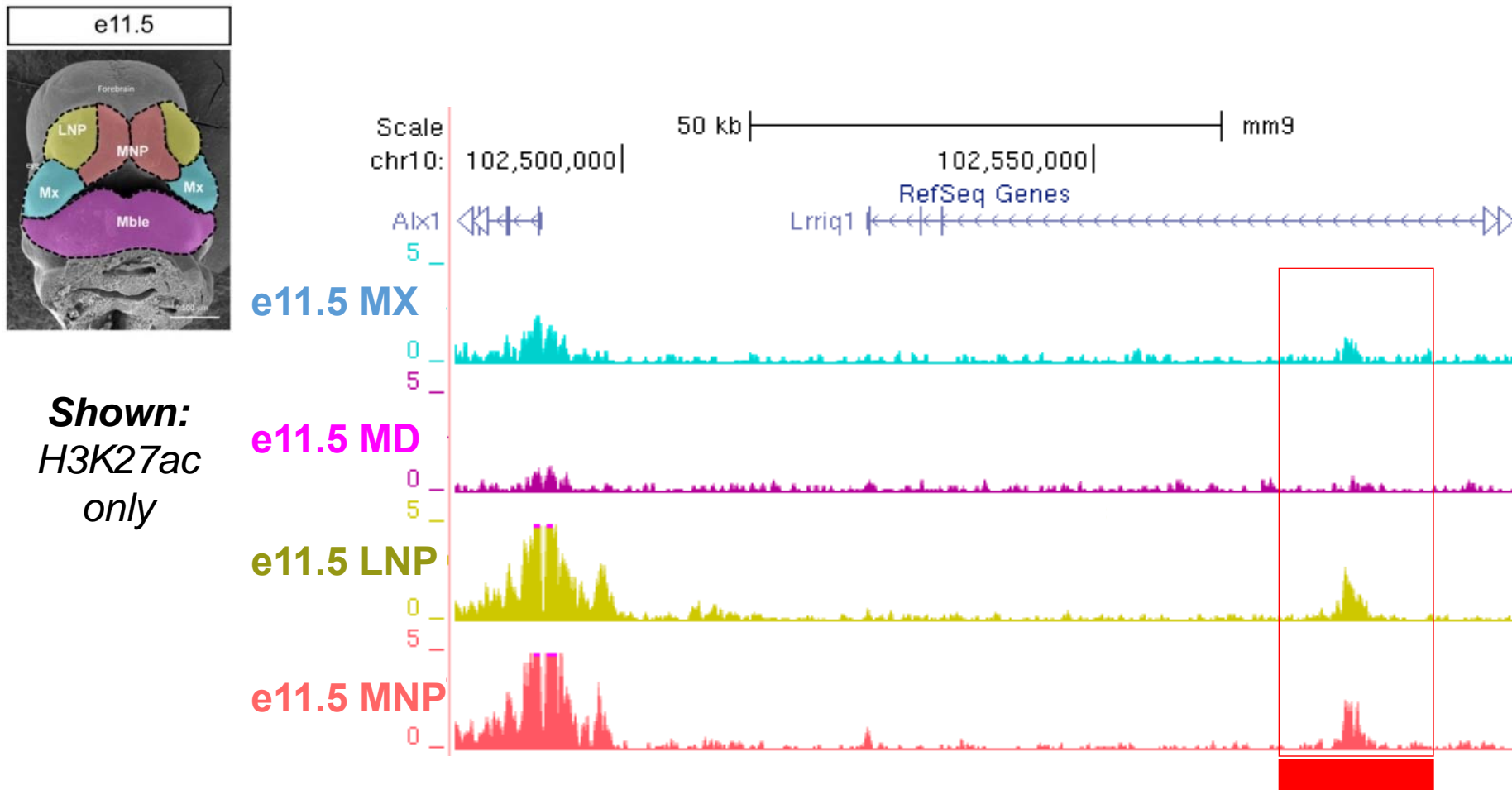
# **Analysis, Interactions and Collaborations**



# Integrative analysis identifies MX/MD-specific enhancer near *Prrx1*



## Integrative analysis identifies NP-specific enhancer near *A/x1*



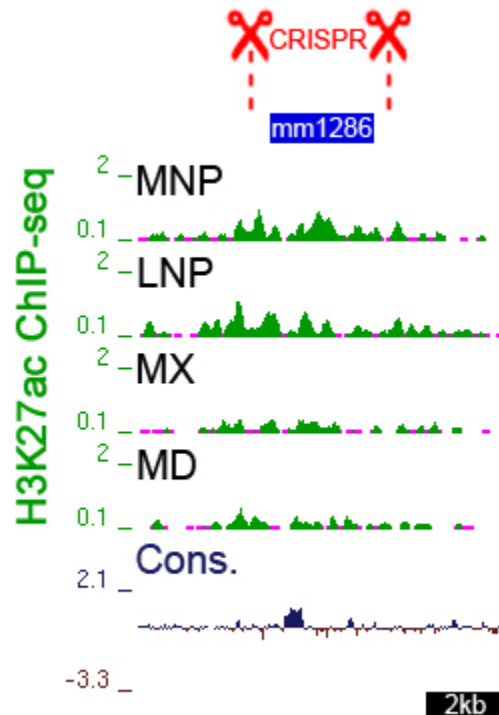
## Larger transgenic series in progress!

*In vivo* LacZ pattern  
Reproducibility: 7/8

# Collaborations and Community Resources

(Example: Enhancer Landscape of GHRL2 - with Seb Dworkin)

***Grhl2* -34kb**

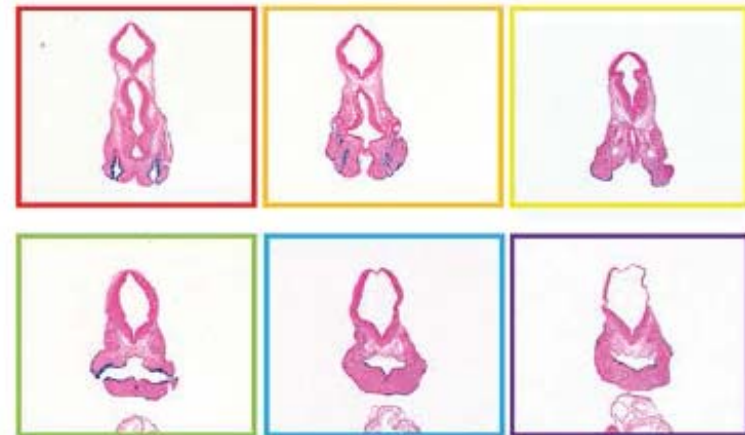
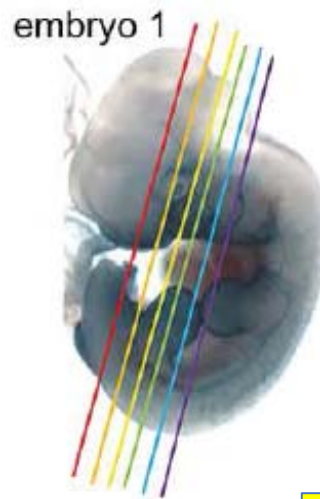


**E11.5**



***Grhl2*** has essential roles in epithelial morphogenesis and epidermal development.

**The mm1286 enhancer is active in craniofacial epithelia at E11.5.**



**Sebastian Dworkin**

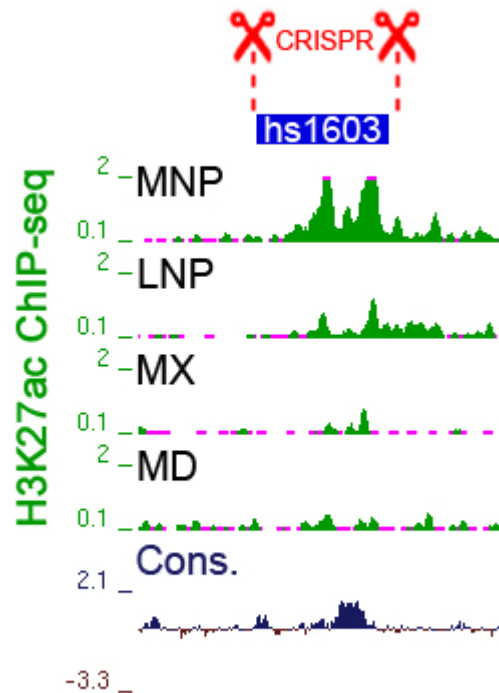
La Trobe University, Melbourne

Enhancer KO line generated and transferred to **Dworkin Lab** for detailed molecular and phenotypic analysis

# Enhancer Function in Known CF Loci

(Example: SOX5 - with Benedikt Hallgrimsson)

**Sox5 +710kb**



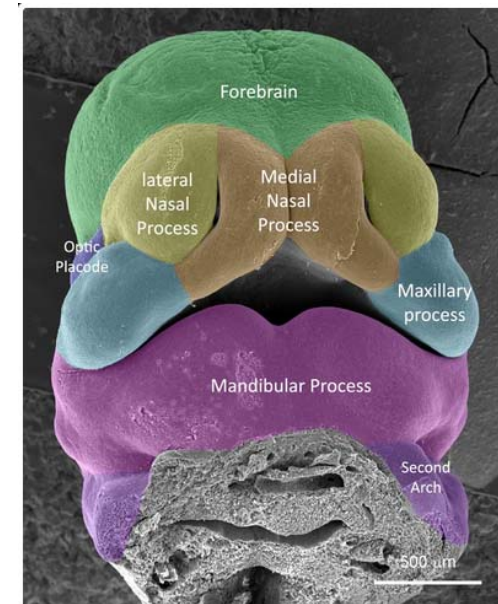
**E11.5**

**MNP**



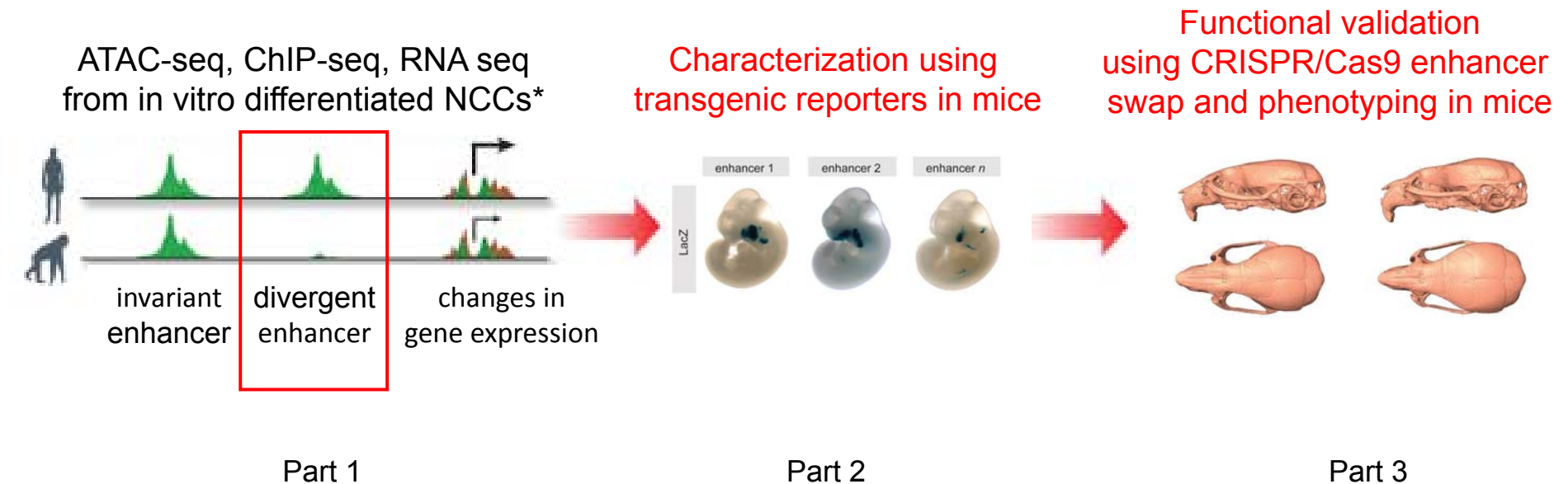
>40 skulls from Enhancer KO and WT controls at p56 shipped to **Hallgrimsson lab**, micro-CT analysis of results in progress

Also: LNP/MNP enhancer near *Bmp4*



LAMB-SHAFFER SYNDROME (OMIM: 616803): associated with **Sox5**;  
→ facial dysmorphologies (e.g. **nasal bridge defects**)












# ***In vivo* Functional Characterization and Validation of Enhancers Driving Morphological Difference Between Human and Chimp Face**



***With Wysocka/Selleri spoke project***



# Validation of Enhancers with Differential Activity

Name	Human	Chimp	Mouse	Human	Putative Target Gene
JW1				Limb (9/10), Periocular Mesenchyme (4/10)	<i>Prrx1/Prx1/MHox</i>
JW3				Trigeminal (6/8), Telencephalon (8/8)	<i>Edn3</i>
JW5				Face, Heart, Somites/DRG, and Otic vesicle (6/7)	<i>Otos</i>
JW6			No blues; 6tg by PCR	Heart, Facial mesenchyme, Hindbrain, Mesenchyme	<i>Mylip</i>

*Knock-in  
in progress*

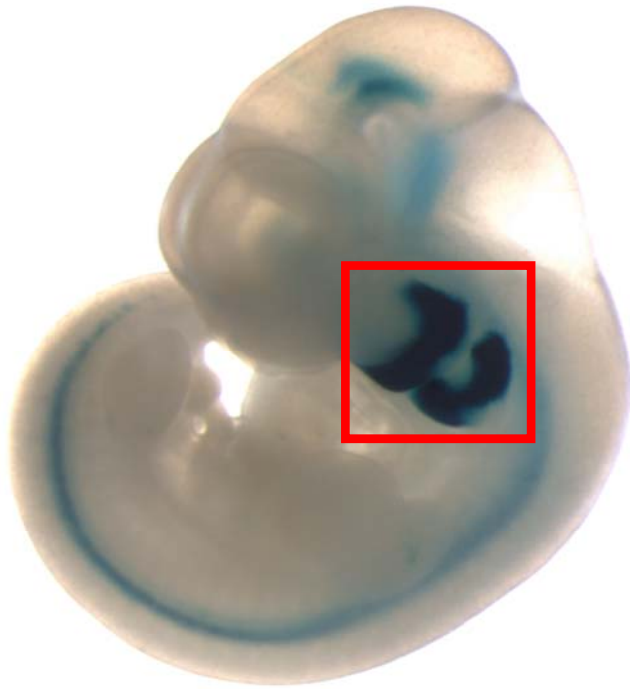
**Also: Sox9 – see Licia's presentation**

**With Wysocka/Selleri spoke project**

# Mapping Enhancers at Single Cell Resolution

# What Cell Type is This Enhancer Active In?

...But typically don't have cell type information



ChIP-Seq, RNA-Seq:

Resolution historically limited to bulk tissue

Transgenic assay:

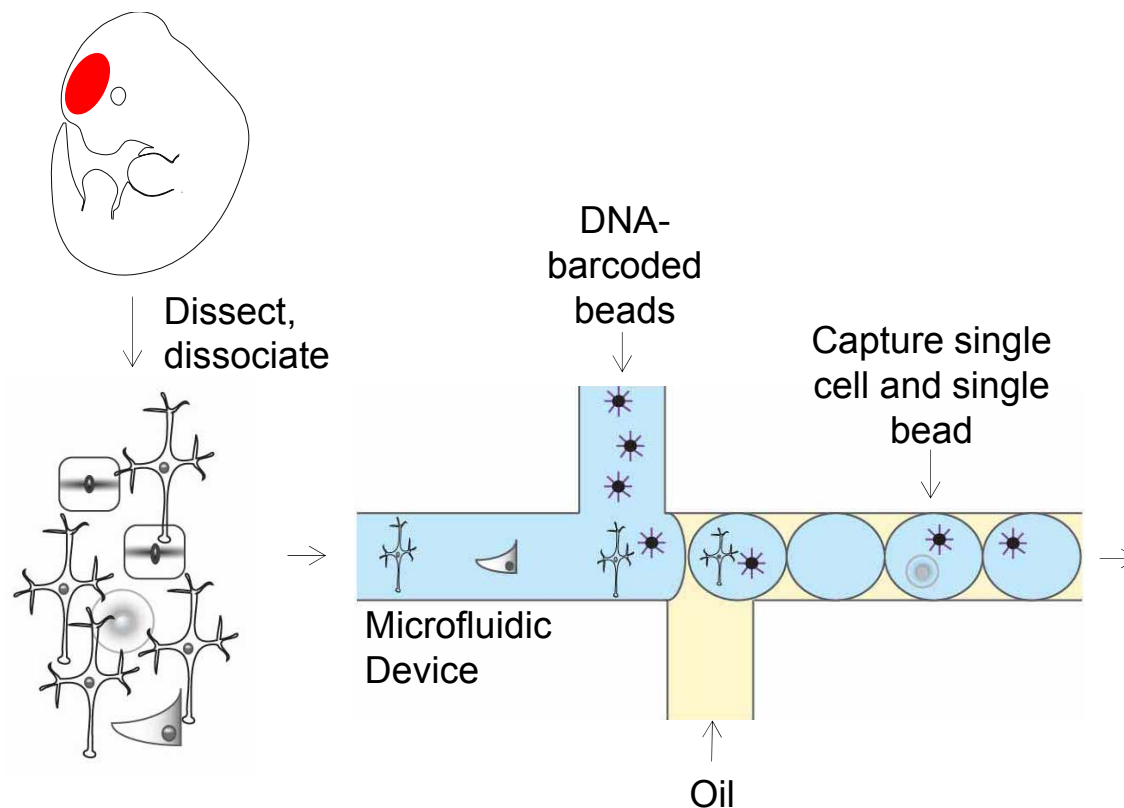
Determining cell type requires laborious immunofluorescence labeling

Enhancers likely define specific cell populations

Can emerging single cell technologies solve this problem?

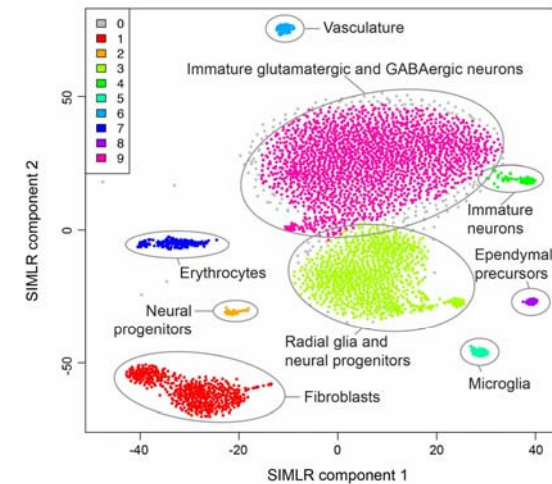


# Single Cell RNA-seq is Becoming Routine



e.g., Drop-seq, 10X Chromium

RNA-seq on >4,700 single brain cells



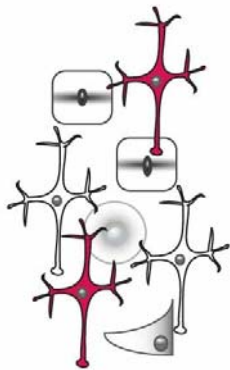
# Single Cell RNA-seq is Becoming Routine

Enhancer active in **forebrain**

hs121 mCherry



Dissect,  
dissociate



Microfluidic  
Device

DNA-  
barcoded  
beads

Capture single  
cell and single  
bead

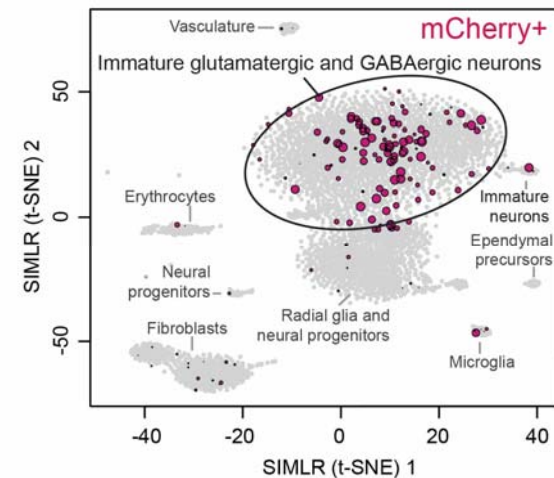
Oil

e.g., Drop-seq, 10X Chromium

Enhancer active in

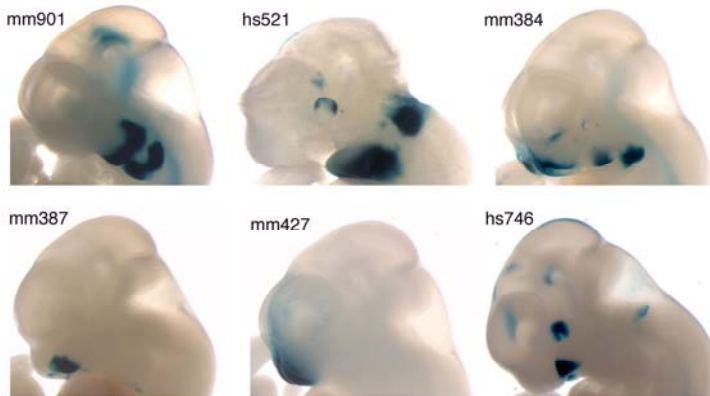
**forebrain immature  
glutamatergic/GABAergic  
neurons**

**Mapping mCherry onto  
RNA-seq on >4,700  
single brain cells**

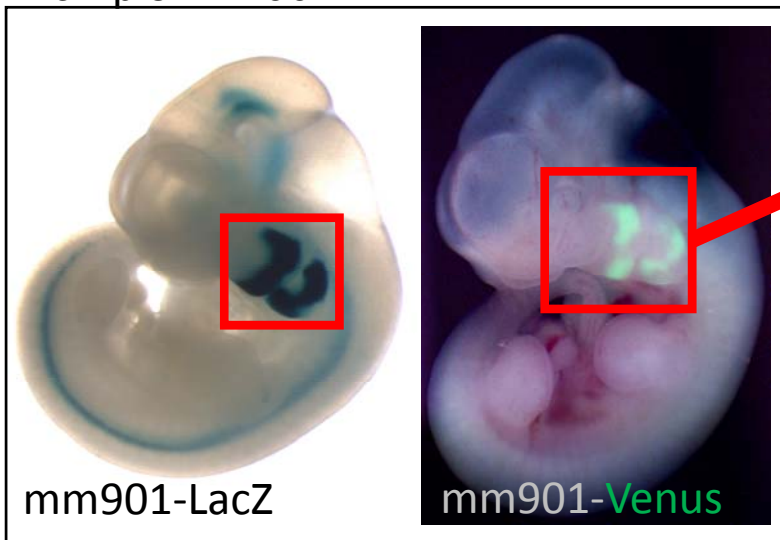


# Single Cell Profiling of E11.5 Face: 28,000+ Cells

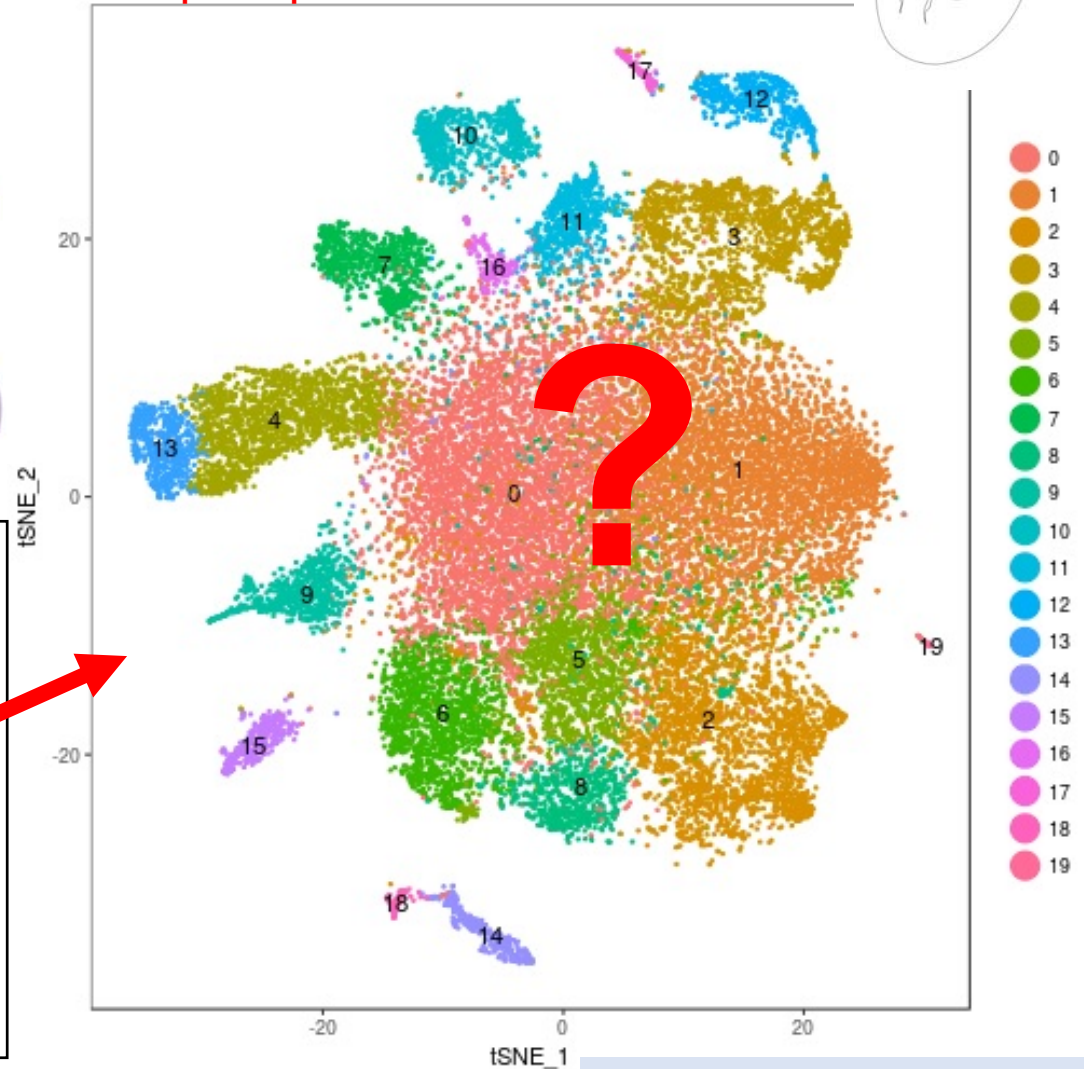
Pilot Study: Six Craniofacial Enhancers



Example: mm901



Drop-seq



Visel Lab, unpublished

# **FaceBase Standardized Analysis Pipelines for ChIP-seq and RNA-seq data**

# FaceBase Standardized Analysis

## Challenge:

- Several FaceBase spokes produce ChIP-seq and/or RNA-seq data
- Each lab has been using their favorite analysis pipeline (in most cases in-house developed/tweaked)
- Limits comparison across data from different spokes and hurdle to integrative analysis

## Goal:

Need to analyze data in a uniform and reproducible way

## Solution:

Analyze data using ENCODE pipelines on DNAnexus

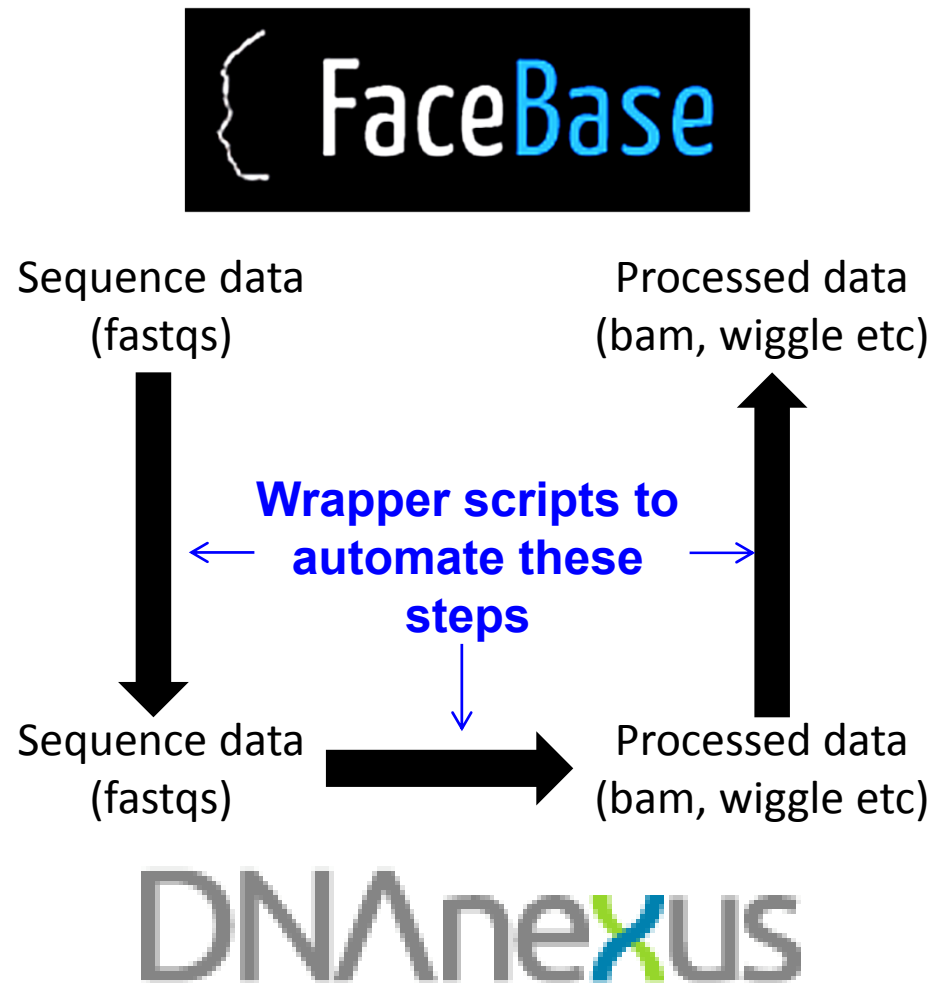
# Using ENCODE Pipelines at DNAnexus

- Platform for genomics data analysis
- Built on Amazon cloud
- Implements analysis tools and workflows including ENCODE pipelines
- Command line interface for automation

DNAnexus

<https://www.dnanexus.com>

# General FaceBase / DNAnexus analysis workflow



# Status of Pipeline Development

## Complete:

- Metadata requirements established and implemented
- Data transfer protocols established
- (Most) wrapper scripts established
- Test data sets successfully processed

## In progress:

- Additional validation and comparison with in-house analyses and replicated data sets
- Streamlining of data transfer (currently temporarily stored at LBNL)
- Ensure human data is properly controlled
- Review display of data in FaceBase



# Project Team

## Lawrence Berkeley National Lab

Experimental postdocs: Sudha Rajderkar, Cailyn Spurrell, Marco Osterwalder, Evgeny Kvon,

Computational postdocs: Valentina Snetkova, Guy Kelman, Iros Barozzi

Other Senior Staff: Diane Dickel, Len Pennacchio

Molecular Biology and Mouse Transgenics: Jennifer Akiyama, Veena Afzal, Brandon Mannion, Cathy Pickle, Ingrid Plaijzer-Frick, Momoe Kato, Tyler Garvin, Elizabeth Lee, Stella Tran

**MRC Human Genetics, Edinburgh, UK - *Optical projection tomography***

**David FitzPatrick, Harris Morrison**

**HDBR, Newcastle, UK**

**Steven Lisgo**

**University of Calgary, Canada - *Morphometry***

**Benedikt Hallgrimsson, Denise Liberton**

**University of Southern California – *KO analysis***

**Yang Chai**

Major Spoke Collaborator:  
Wysocka Lab

